



Convenient synthesis of C-terminal di- and tri-peptide amides from *N*-protected dipeptidoylbenzotriazoles

Ilhami Celik^a, Ashraf A.A. Abdel-Fattah^{b,*}

^a Department of Chemistry, Faculty of Science, Anadolu University, 26470 Eskisehir, Turkey

^b Department of Chemistry, Faculty of Science, Benha University, Benha, Egypt

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ABSTRACT

N-Protected dipeptidoylbenzotriazoles react with aqueous ammonia to give dipeptide primary amides (77–98%) and with *N*-unprotected α -amino amides to afford tripeptide primary amides (82–86%).

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1. Introduction

Naturally occurring peptides are essential for many cellular functions.¹ Peptides modified at the C-terminus are substrates or inhibitors for a variety of proteolytic enzymes.² The incorporation of a primary amide at the C-terminus is common modification:³ around 50% of biologically active peptides contain such functionality.⁴ Peptidyl α -carboxamides are especially widely distributed in animals and elicit vital physiological effects:^{1a} examples include oxytocin,⁵ luteinizing hormone releasing hormone (LHRH),⁶ neurokinins,⁷ and vasopressin and gastrin.⁸

Peptidyl α -carboxamides are also found within protease inhibitors. For example, the pseudo pentapeptide 1-682,679 (**1**) is a highly potent and selective inhibitor of HIV-1 protease (IC_{50} =0.42 nM, Fig. 1) (**11**) and 1-685,458 (**2**) is a potent inhibitor of γ -secretase (IC_{50} =17 nM) of potential therapeutic benefit in the treatment of Alzheimer's disease and other neurological disorders (Fig. 1).⁹ Cathepsin L (EC 3.4.22.16) is an example of the papain-like cysteine proteases, which are ubiquitously distributed in the lysosomes of cells with prominent roles in nonspecific intracellular protein breakdown,¹⁰ and physiologically important extra-cellular functions.¹¹

Recently, an *N*^ω-nitroarginine-containing dipeptide amide [*L*-Arg^{NO2}-*L*-Lys-NH₂ (**LL**) (**3**)] and [*D*-Lys-*D*-Arg^{NO2}-NH₂ (**DD**) (**4**)] (Fig. 2)¹² showed a remarkable selectivity (hundreds- to thousands-fold) for inhibition of the neuronal isozyme of nitric oxide synthase (*n*NOS) over two other isozymes.¹³ This *n*NOS selectivity has implications in the potential treatment of strokes,¹⁴ migraine headaches,¹⁵ and Alzheimer's disease.¹⁶ Moreover, *L*-Leu-*x*-amides

(where *x* is any amino acid residue) support growth of a leucine auxotroph as well as the free peptide.¹⁷

N-Acylbenzotriazoles are neutral efficient coupling reagents for *N*-acylation,¹⁸ *C*-acylation,¹⁹ and *O*-acylation.²⁰ *N*-(Aminoacyl)-benzotriazoles prepared from *N*-protected α -amino acids were successfully utilized for synthesis of di-, tri-, and tetrapeptides.²¹ As part of the synthetic strategy for the modification of peptides at the C-terminus, we now report the synthesis of C-terminal protected dipeptides **10a–l**, (**10a+10a'**),²² (**10e+10e'**) and tripeptide amides **13a,b** and (**13a+13a'**) by treating *N*-protected dipeptidoylbenzotriazoles **9a–l**, (**9a+9a'**), and (**9e+9e'**) with aqueous ammonia or with unprotected α -amino carboxamides **12a,b**, respectively.

2. Results

2.1. Synthesis of *N*-protected dipeptidoylbenzotriazoles

Treating Cbz-protected amino acids **5a–e** and (**5b'**) with 4 equiv of benzotriazole and 1 equiv of SOCl₂ in THF or CH₂Cl₂ at 20 °C for 2 h, following the published procedure,^{21a,c} gave *N*-(Cbz- α -aminoacyl)benzotriazoles **6a–e** and (**6b'**) in 78–95% (Scheme 1). Peptide coupling was achieved by the reaction of **6a–e** and (**6b'**) with unprotected amino acids **7a–g** and (**7a'**) in aqueous acetonitrile in the presence of Et₃N for 10–40 min.^{21b} After washing with 6 M HCl, the resulting dipeptides **8a–l**, (**8a+8a'**), and (**8e+8e'**) were obtained in 71–95% yields (Scheme 2 and Table 1). These reactions proceed without detectable racemization of the chiral center, as evidenced by their NMR and HPLC analysis.^{21a,c,e}

Cbz-protected dipeptidoylbenzotriazoles **9a–l**, (**9a+9a'**), and (**9e+9e'**) were prepared in 64–91% yield from Cbz-protected dipeptides **8a–l**, (**8a+8a'**), and (**8e+8e'**) by reaction with 4 equiv of benzotriazole and 1 equiv of SOCl₂ at –15 °C for 4–6 h (Scheme 2 and Table 2).^{21b–d}

* Corresponding author.

E-mail address: ashraf25894@yahoo.com (A.A.A. Abdel-Fattah).

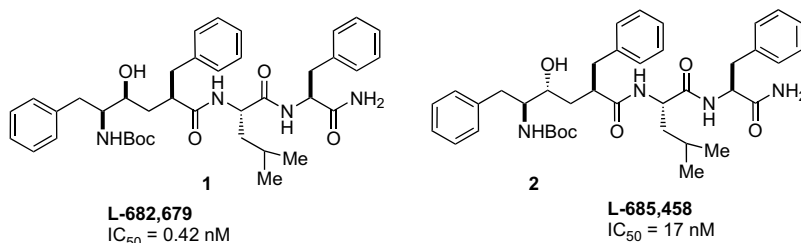


Figure 1. Chemical structures of the aspartyl protease inhibitors L-682,679 and L-685,458.

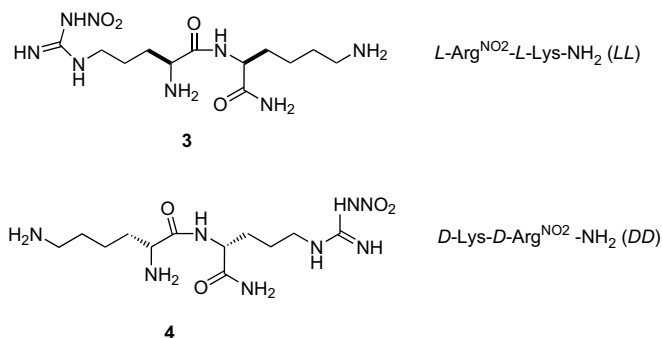


Figure 2. Chemical structures of L-Arg^{NO2}-L-Lys-NH₂ (LL) and D-Lys-D-Arg^{NO2}-NH₂ (DD).

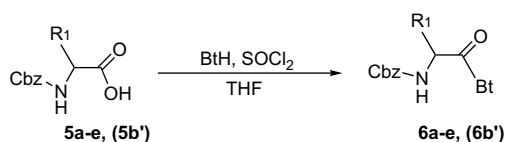
2.2. Synthesis of *N*-protected dipeptide primary amides

Treatment of the *N*-protected dipeptidoylbenzotriazoles **9a–l**, (**9a+9a'**), and (**9e+9e'**) with excess aqueous ammonium hydroxide in THF at 0 °C for 2 h afforded the corresponding dipeptide primary amides **10a–l**, (**10a+10a'**), and (**10e+10e'**) in 77–98% yield (Scheme 2 and Table 3). The structures of **10a–l**, (**10a+10a'**), and (**10e+10e'**) were supported by spectral data and elemental analyses.

NMR analysis showed no detectable racemization for compounds **10a–l**. No signals from their corresponding diastereomers were observed in the NMR spectra of **10a–l** suggesting the enantiopurity of the *N*-protected dipeptide amides. For example, the ¹H NMR spectrum of **10a** gave a clear doublet for the methyl protons of L-Ala fragment at 1.19 ppm, whereas a multiplet was observed for the corresponding diastereomeric mixture (**10a+10a'**) ranging from 1.08 to 1.23 ppm. Furthermore, compound **10a** and the corresponding diastereomeric mixture (**10a+10a'**) were subjected to HPLC analysis using Chirobiotic T column (detection at 254 nm, flow rate 1.0 mL/min, and 100% MeOH as solvent). As expected, HPLC analysis of the enantiopure L-dipeptide amide **10a** showed a signal peak at 3.86 min. In contrast, two peaks of equal intensity at 3.67 and 3.79 min were observed for the corresponding diastereomeric mixture (**10a+10a'**).

2.3. Synthesis of *N*-protected tripeptide primary amides

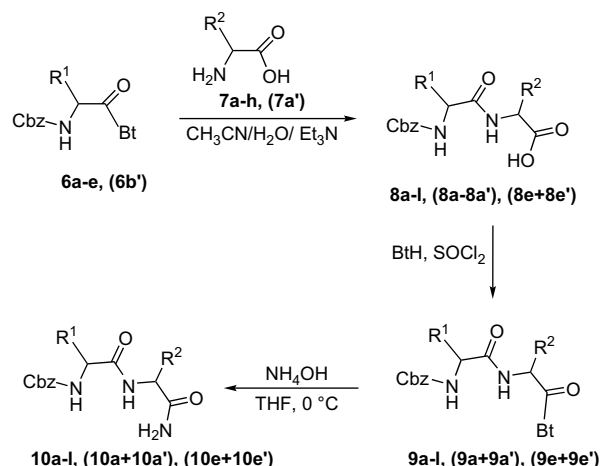
Synthesis of *N*-protected tripeptide primary amides **13a,b** and (**13a+13a'**) was achieved by reacting the *N*-protected



Bt = Benzotriazol-1-yl

R¹: a, L-Trp; b, L-Ala; b', DL-Ala; c, L-Met; d, L-Phe; e, D-Val

Scheme 1. Compound numbers written within brackets represent a diastereomeric mixtures or racemates; compound numbers without brackets represent enantiomers.



Scheme 2.

dipeptidoylbenzotriazoles **9a,b** and (**9a+9a'**) (described above) with *N*-unprotected-α-amino amides **12a,b** (prepared by treating the corresponding *N*-Fmoc-α-aminoacylbenzotriazoles **11a** (Fmoc-L-Met-Bt) and **11b** (Fmoc-Gly-Bt) with aqueous ammonia followed by *N*-deprotection with piperidine utilizing a literature method²³ in acetonitrile at –15 °C for 6 h to afford the corresponding *N*-protected tripeptide amides **13a,b** and (**13a+13a'**) in 82–86% yields (Scheme 3 and Table 4).

NMR spectroscopic analysis of the tripeptide amide **13a** showed approximately 25% racemization. The ¹H NMR spectrum of **13a** displayed two separate doublets with different intensities for the methyl protons of L-Ala fragment at 1.13 ppm and 1.26 ppm,

Table 1
Preparation of dipeptides **8**

<i>N</i> -(Cbz-α-aminoacyl) benzotriazoles	Amino acid	Product	Yield ^a (%)
6a	L-Ala, 7a	Cbz-L-Trp-L-Ala-OH, 8a	95
6a	DL-Ala, (7a')	Cbz-L-Trp-DL-Ala-OH, (8a+8a')	93
6a	L-Trp, 7b	Cbz-L-Trp-L-Trp-OH, 8b	86
6a	Gly, 7c	Cbz-L-Trp-Gly-OH, 8c	93
6b	L-Trp, 7d	Cbz-L-Ala-L-Trp-OH, 8d	79
6b	L-Glu(OBn), 7e	Cbz-L-Ala-L-Glu(OBn)-OH, 8e	91
6b'	L-Glu(OBn), 7e	Cbz-DL-Ala-L-Glu(OBn)-OH, (8e+8e')	81
6b	L-Pro, 7f	Cbz-L-Ala-L-Pro-OH, 8f	78
6c	L-Trp, 7d	Cbz-L-Met-L-Trp-OH, 8g	86
6d	L-Met, 7f	Cbz-L-Phe-L-Met-OH, 8h	70
6d	L-Glu(OBn), 7e	Cbz-L-Phe-L-Glu(OBn)-OH, 8i	93
6d	L-Asp(OBn), 7g	Cbz-L-Phe-L-Asp(OBn)-OH, 8j	79
6d	L-Lys, 7h	Cbz-L-Phe-L-Lys-OH, 8k	71
6f	L-Asp(OBn), 7g	Cbz-D-Val-L-Asp(OBn)-OH, ^b 8l	83

^a Isolated yield.

^b Novel compound.

Table 2
Conversion of dipeptides **8** into *N*-Cbz-dipeptidoylbenzotriazoles **9**

Reactant	Product	Yield ^a (%)
8a	Cbz-L-Trp-L-Ala-Bt, 9a	79
(8a+8a')	Cbz-L-Trp-DL-Ala-Bt, (9a+9a') ^b	64
8b	Cbz-L-Trp-L-Trp-Bt, 9b	89
8c	Cbz-L-Trp-Gly-Bt, 9c ^b	75
8d	Cbz-L-Ala-L-Trp-Bt, 9d	78
8e	Cbz-L-Ala-L-Glu(OBn)-Bt, 9e	91
(8e+8e')	Cbz-DL-Ala-L-Glu(OBn)-Bt, (9e+9e')	79
8f	Cbz-L-Ala-L-Pro-Bt, 9f ^b	76
8g	Cbz-L-Met-L-Trp-Bt, 9g ^b	73
8h	Cbz-L-Phe-L-Met-Bt, 9h	79
8i	Cbz-L-Phe-L-Glu(OBn)-Bt, 9i	80
8j	Cbz-L-Phe-L-Asp(OBn)-Bt, 9j	81
8k	Cbz-L-Phe-L-Lys-Bt, 9k	63
8l	Cbz-D-Val-L-Asp(OBn)-Bt, 9l ^b	88

^a Isolated yield.^b Novel compound.

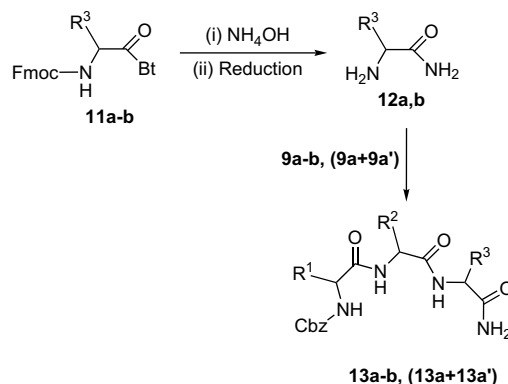
respectively. However for compound **13b** no signals for any corresponding diastereomer were observed. The enantiopurity of tripeptide amide **13b** was further confirmed by HPLC analyses using Chirobiotic T column (detection at 254 nm, flow rate 1 mL/min, and MeOH as solvent): tripeptide amide **13b** gave a single retention time, whereas the diastereomeric mixture (**13a+13a'**) showed two retention times as expected (Table 4).

Traditional syntheses of primary carboxamides (i) treat acids or derived acid halides, anhydrides or esters with ammonia.²⁴ However, the exothermic reactions of ammonia with acyl halides must be controlled; acid anhydrides and ammonia can also form imides.²⁵ Additionally, these methods are often not suitable for selective amidation.²⁶ Other reported methods include: (ii) ammonolysis of *N*-protected amino acids with diaminomethane dihydrochloride,²⁷ (iii) treatment of acids with ammonia-releasing agents (NH₄OH, NH₄Cl or (NH₄)₂CO₃) in the presence of benzotriazol-1-yl-oxytripyrrolidinophosphonium hexafluoro-phosphate (PyBOP) and *N*-hydroxybenzotriazole (HOBt),²⁸ 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) and *N*-hydroxy-7-aza-benzotriazole (HOAt) or HOBt (**48**), *N,N'*-dicyclohexyl-carbodiimide (DCC) and HOAt or HOBt,²⁹ *p*-tolylsulfonyl chloride (TsCl),³⁰ or *N*-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ).³¹ Difficulties can arise from the insolubility of starting materials, competitive hydrolysis or partial racemization in approaches (ii) and (iii).

Peptidyl α -carboxamides have also been prepared previously by several solid phase and biosynthetic methods. Solid phase syntheses include (iv) acidolytic cleavage of peptide from support-bound benzylamine- or benzhydrylamine-based handles,³² (but

Table 3
Conversion of **9** into dipeptide amides **10**

Reactant	Product	Yield ^a (%)	Mp (°C)	[α] _D ²³
9a	Cbz-L-Trp-L-Ala-NH ₂ , 10a ^b	79	168–170	–9.5
(9a+9a')	Cbz-L-Trp-DL-Ala-NH ₂ , (10a+10a') ^c	95	157–160	–20.7
9b	Cbz-L-Trp-L-Trp-NH ₂ , 10b	96	192–194	–36.6
9c	Cbz-L-Trp-Gly-NH ₂ , 10c	86	151–155	–26.4
9d	Cbz-L-Ala-L-Trp-NH ₂ , 10d	77	201–202	–7.7
9e	Cbz-L-Ala-L-Glu(OBn)-NH ₂ , 10e	98	162–163	–3.0
(9e+9e')	Cbz-DL-Ala-L-Glu(OBn)-NH ₂ , (10e+10e')	90	138–139	–4.7
9f	Cbz-L-Ala-L-Pro-NH ₂ , 10f	95	161–162	–37.6
9g	Cbz-L-Met-L-Trp-NH ₂ , 10g	91	218–220	–12.7
9h	Cbz-L-Phe-L-Met-NH ₂ , 10h	94	208–210	–14.8
9i	Cbz-L-Phe-L-Glu(OBn)-NH ₂ , 10i	96	205–207	–10.6
9j	Cbz-L-Phe-L-Asp(OBn)-NH ₂ , 10j	98	162–163	–3.0
9k	Cbz-L-Phe-L-Lys-NH ₂ , 10k	92	193–195	–8.5
9l	Cbz-D-Val-L-Asp(OBn)-NH ₂ , 10l	88	195–196	–32.9

^a Isolated yield.^b HPLC for **10a**: 3.86 min.^c HPLC for (**10a+10a'**): 3.67 and 3.79 min.**Scheme 3.**

side-chain deprotection takes place in concert with cleavage); (v) ammonolysis of support-bound esters of side-chain deprotected peptide with liquid ammonia or ammonia/MeOH solution,³³ or ammonia/THF,³⁴ (limited to side-chain deprotected peptides and requiring well-sealed containers, as trace amounts of moisture can result in the production of acid as a contaminant);³⁵ and (vi) deprotection and cleavage of solid-supported *N*-tetrachlorophthaloyl peptides.³⁶

Biosynthetic techniques include: (vii) oxidative cleavage of polypeptides by peptidylglycine α -amidating monooxygenase (PAM);³⁷ (viii) enzymatic couplings of an acyl donor component (esters or thioesters) with an acyl acceptor component (amino acid amides).³⁸ Enzymatic approaches have attracted interest for years because they take place in an aqueous medium and occur with high stereo- and regio-selectivity in mild conditions of pH and temperature with no need for side-chain protection. However, they may be restricted to specific substrates, require long reaction times, and/or give the products in low yield.³⁸

We have shown that our methodology offers convenient preparative and work-up procedures, and uses inexpensive reagents to prepare *N*-protected di- and tripeptide primary amides from readily available *N*-protected dipeptidoylbenzotriazoles in good yields (77–98%) as compared with literature methods.

3. Summary

In summary, we have developed a convenient and efficient approach for a broad range of chirally pure dipeptide primary amides (average yield 88%), by treating the corresponding *N*-protected dipeptidoylbenzotriazoles with aqueous ammonia. The approach was extended to the preparation of tripeptide primary amides (average yield 83%) by using some of the same substrates with *N*-unprotected α -amino amides. These results, coupled with the simplicity of isolation and purification procedures, make this approach competitive with literature methods.

Table 4
Conversion of **9a,b** and **(9a+9a')** into tripeptide amides **13**

Reactant	Amino amide	Product	Yield ^a (%)	Mp (°C)	[α] _D ²³
9a	Met-NH ₂ , 12a	Cbz-L-Trp-L-Ala-Met-NH ₂ , 13a	82	222–224	–33.30
(9a+9a')	Gly-NH ₂ , 12b	Cbz-L-Trp-DL-Ala-Gly-NH ₂ , (13a+13a')	82	164–166	–17.92
9b	Gly-NH ₂ , 12b	Cbz-L-Trp-L-Trp-Gly-NH ₂ , 13b	86	220–221	–15.36

^a Isolated yield.^b HPLC for (**13a+13a'**): 9.90 and 11.82 min.^c HPLC for **13b**: 8.17 min.

4. Experimental

4.1. General

All mps are uncorrected. NMR spectra were recorded on a Varian Gemini (300 MHz) spectrometer in DMSO- d_6 with TMS for ^1H (300 MHz) and ^{13}C (75 MHz) as an internal reference. IR spectra were recorded on a Unicam SP-1200 infrared spectrophotometer using KBr Wafer technique. *N*-Cbz-amino acids and amino acids were purchased from Fluka (Buchs, Switzerland) and Acros (Suwanee, GA, USA), were used without further purification. Elemental analyses were performed on a Carlo Erba-1106 instrument. Optical rotation values were measured with the use of the sodium D line. HPLC analyses were performed on Beckman system gold programmable solvent module 126 using Chirobiotic T column (4.6×250 mm), detection at 254 nm, flow rate 1.0 mL/min, and with methanol as solvent.

4.2. *N*-(Cbz- α -aminoacyl)benzotriazoles **6**: general procedure

N-(Cbz- α -aminoacyl)benzotriazoles **6a–e** and (**6b'**) were prepared according to the literature procedure.^{18,20,21a}

4.3. Dipeptides **8**: general procedure

N-Cbz-protected dipeptides **8a–l**, (**8a+8a'**), and (**8e+8e'**) were prepared according to the literature procedure.^{38,20,21a}

4.3.1. (3*S*)-4-(Benzyloxy)-3-[(2*R*)-2-[(benzyloxy)carbonyl]amino-3-methylbutanoyl]amino]-4-oxobutanoic acid (Cbz-*D*-Val-*L*-Asp(3-OBn)-OH, **8l**)

Yield: 83%; white microcrystals; mp 114–115 °C; $[\alpha]_D^{23}$ –19.3 (c 1.65, DMF). ^1H NMR: δ =8.42 (d, J =7.8 Hz, 1H), 7.39–7.30 (m, 11H), 5.12 (s, 2H), 5.07 (s, 2H), 4.65 (dd, J =13.5, 7.0 Hz, 1H), 3.96 (t, J =7.8 Hz, 1H), 2.91 (dd, J =16.3, 5.6 Hz, 1H), 2.75 (dd, J =16.4, 7.2 Hz, 1H), 1.98–1.92 (m, 1H), 0.84 (t, J =6.5 Hz, 6H). ^{13}C NMR: δ =172.1, 171.2, 170.2, 156.3, 137.3, 136.1, 128.6, 128.5, 128.2, 128.0, 127.9, 127.8, 65.9, 65.5, 60.0, 48.7, 36.1, 30.7, 19.2, 18.0. IR: ν_{max} 3430–2500 (br), 1705, 1645, 1620, 1590, 1425 cm^{-1} . Anal. Calcd for $\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_7$: C, 63.15; H, 6.18; N, 6.14. Found: C, 62.82; H, 6.34; N, 6.29.

4.4. *N*-Cbz-protected dipeptidoylbenzotriazoles **9**: general procedure

N-Cbz-protected dipeptidoylbenzotriazoles **9a–l**, (**9a+9a'**), and (**9e+9e'**) were prepared according to the literature procedure.^{18,21}

4.4.1. Benzyl *N*-[(1*S*)-2-[2-(1*H*-1,2,3-benzotriazol-1-yl)-1-methyl-2-oxoethyl]amino-1-(1*H*-indol-2-ylmethyl)-2-oxoethyl]carbamate (Cbz-*L*-Trp-*D*-Ala-*Bt*, (**9a+9a'**))

Yield: 64%; white microcrystals; mp 142–144 °C; $[\alpha]_D^{23}$ –8.4 (c 1.81, DMF). ^1H NMR: δ =10.84 (s, 1H), 9.04–8.97 (m, 1H), 8.32–8.24 (m, 2H), 7.82 (t, J =7.7 Hz, 1H), 7.73–7.63 (m, 2H), 7.44 (d, J =8.7 Hz, 1H), 7.35–7.19 (m, 7H), 7.06–6.97 (m, 2H), 5.67–5.63 (m, 1H), 4.93 (d, J =3.7 Hz, 2H), 4.45–4.41 (m, 1H), 3.16–3.12 (m, 1H), 2.94–2.91 (m, 1H), 1.61–1.49 (m, 3H). ^{13}C NMR: δ =172.6, 172.3, 171.9, 155.9, 155.7, 145.4, 137.0, 136.1, 131.1, 130.7, 128.3, 127.7, 127.5, 127.4, 127.2, 126.9, 126.7, 124.1, 124.0, 120.8, 120.2, 118.6, 118.2, 114.0, 111.3, 110.0, 109.9, 65.3, 65.0, 55.1, 54.9, 48.7, 48.6, 31.0, 27.8, 16.7, 16.6. IR: ν_{max} 3100, 3060, 2915, 1715, 1650, 1620, 1410 cm^{-1} . Anal. Calcd for $\text{C}_{28}\text{H}_{26}\text{N}_6\text{O}_4$: C, 65.87; H, 5.13; N, 16.46. Found: C, 65.61; H, 5.27; N, 16.43.

4.4.2. Benzyl *N*-[(1*S*)-2-[2-(1*H*-1,2,3-benzotriazol-1-yl)-2-oxoethyl]amino-1-(1*H*-indol-2-ylmethyl)-2-oxoethyl]carbamate (Z-*L*-Trp-*L*-Gly-*Bt*, **9c**)

Yield: 75%; white microcrystals; mp 186–187 °C; $[\alpha]_D^{23}$ –27.3 (c 1.73, DMF). ^1H NMR: δ =10.87 (s, 1H), 8.89 (t, J =5.1 Hz, 1H), 8.30 (d,

J =8.2 Hz, 1H), 8.25 (d, J =8.2 Hz, 1H), 7.82 (t, J =7.6 Hz, 1H), 7.72 (d, J =7.7 Hz, 1H), 7.64 (t, J =7.7 Hz, 1H), 7.56 (d, J =8.5 Hz, 1H), 7.37–7.23 (m, 7H), 7.08 (t, J =7.3 Hz, 1H), 7.00 (t, J =7.3 Hz, 1H), 5.02 (t, J =5.9 Hz, 2H), 4.97 (s, 2H), 4.49–4.44 (m, 1H), 3.25 (dd, J =14.4, 3.4 Hz, 1H), 3.00 (dd, J =14.4, 10.5 Hz, 1H). ^{13}C NMR: δ =173.0, 169.6, 155.9, 145.3, 137.0, 131.0, 130.6, 128.3, 127.7, 127.5, 127.2, 126.6, 124.0, 120.9, 120.2, 118.5, 118.2, 113.8, 111.3, 110.2, 65.3, 55.4, 42.6, 27.9. Anal. Calcd for $\text{C}_{27}\text{H}_{24}\text{N}_6\text{O}_4$: C, 65.31; H, 4.87; N, 16.93. Found: C, 65.63; H, 4.99; N, 16.55.

4.4.3. Benzyl *N*-[(1*S*)-2-[(2*S*)-2-(1*H*-1,2,3-benzotriazol-1-yl)carbonyl]tetrahydro-1*H*-pyrrol-1-yl]-1-methyl-2-oxoethylcarbamate (Cbz-*L*-Ala-*L*-Pro-*Bt*, **9f**)

Yield: 76%; white microcrystals; mp 52–54 °C; $[\alpha]_D^{23}$ –29.8 (c 1.66, DMF). ^1H NMR: δ =8.34 (d, J =8.2 Hz, 1H), 8.26 (d, J =8.1 Hz, 1H), 7.84 (t, J =7.7 Hz, 1H), 7.71–7.61 (m, 2H), 7.42–7.33 (m, 5H), 5.76 (dd, J =8.8, 5.1 Hz, 1H), 5.04 (AB, J_{AB} =16.2, 12.6 Hz, 2H), 4.46 (t, J =7.2 Hz, 1H), 3.88–3.76 (m, 2H), 2.50–2.46 (m, 1H), 2.25–2.10 (m, 3H), 1.27 (d, J =7.0 Hz, 3H). ^{13}C NMR: δ =171.4, 170.7, 155.8, 145.5, 137.2, 131.3, 130.8, 128.5, 128.0, 127.9, 127.0, 120.4, 114.1, 65.6, 59.3, 48.1, 47.0, 29.0, 25.3, 16.7. Anal. Calcd for $\text{C}_{22}\text{H}_{23}\text{N}_5\text{O}_4$: C, 62.70; H, 5.50; N, 16.62. Found: C, 62.49; H, 5.57; N, 16.91.

4.4.4. Benzyl *N*-[(1*S*)-1-[(1*S*)-2-(1*H*-1,2,3-benzotriazol-1-yl)-1-(1*H*-indol-2-ylmethyl)-2-oxoethyl]-aminocarbonyl]-3-(methylsulfanyl)propyl]carbamate (Cbz-*L*-Met-*L*-Trp-*Bt*, **9g**)

Yield: 73%; white microcrystals; mp 171–173 °C; $[\alpha]_D^{23}$ +18.0 (c 1.58, DMF). ^1H NMR: δ =10.94 (s, 1H), 8.91 (d, J =5.8 Hz, 1H), 8.29 (d, J =8.1 Hz, 1H), 8.22 (d, J =8.2 Hz, 1H), 7.82 (t, J =7.6 Hz, 1H), 7.66 (t, J =7.6 Hz, 1H), 7.59–7.55 (m, 2H), 7.38–7.28 (m, 7H), 7.06 (t, J =7.5 Hz, 1H), 6.96 (t, J =7.3 Hz, 1H), 5.96–5.90 (m, 1H), 5.04 (AB, J_{AB} =17.3, 12.4 Hz, 2H), 4.28–4.26 (m, 1H), 3.53 (dd, J =14.7, 4.8 Hz, 1H), 3.37–3.31 (m, 1H), 2.51 (t, J =7.8 Hz, 2H), 2.07 (s, 3H), 1.94–1.85 (m, 2H). ^{13}C NMR: δ =172.2, 171.4, 155.9, 145.3, 136.9, 136.0, 131.0, 130.5, 128.3, 127.8, 128.6, 126.7, 124.2, 121.0, 120.2, 118.4, 118.0, 114.0, 111.4, 108.7, 65.7, 53.8, 53.4, 31.7, 29.5, 26.9, 14.6. Anal. Calcd for $\text{C}_{30}\text{H}_{30}\text{N}_6\text{O}_4\text{S}$: C, 63.14; H, 5.30; N, 14.73. Found: C, 62.98; H, 5.21; N, 15.04.

4.4.5. Benzyl (3*S*)-4-(1*H*-1,2,3-benzotriazol-1-yl)-3-[(2*R*)-2-[(benzyloxy)carbonyl]amino-3-methylbutanoyl]amino]-4-oxobutanoate (Cbz-*D*-Val-*Asp*(OBn)-*Bt*, **9l**)

Yield: 86%; white microcrystals; mp 109–110 °C; $[\alpha]_D^{23}$ –10.8 (c 1.80, DMF). ^1H NMR: δ =9.05 (d, J =6.7 Hz, 1H), 8.27 (d, J =8.4 Hz, 1H), 8.21 (d, J =8.2 Hz, 1H), 7.81 (t, J =7.6 Hz, 1H), 7.64 (t, J =7.7 Hz, 1H), 7.36–7.29 (m, 11H), 5.98 (dd, J =13.7, 6.7 Hz, 1H), 5.17–4.98 (m, 4H), 3.97 (dd, J =15.3, 6.7 Hz, 1H), 3.30 (dd, J =16.8, 5.4 Hz, 1H), 3.01 (dd, J =16.8, 8.4 Hz, 1H), 1.96–1.89 (m, 1H), 0.81 (d, J =6.9 Hz, 3H), 0.75 (d, J =6.6 Hz, 3H). ^{13}C NMR: δ =171.8, 169.9, 169.6, 156.3, 145.5, 137.2, 135.8, 131.2, 130.8, 128.5, 128.4, 128.2, 128.1, 128.0, 127.9, 127.8, 126.9, 120.4, 114.1, 66.3, 65.6, 59.9, 49.7, 35.5, 30.6, 19.2, 18.0. Anal. Calcd for $\text{C}_{30}\text{H}_{31}\text{N}_5\text{O}_6$: C, 64.62; H, 5.60; N, 12.56. Found: C, 64.55; H, 5.59; N, 12.28.

4.5. Dipeptide primary amides **10**: general procedure

To a solution of *N*-protected dipeptidoylbenzotriazoles **9a–l**, (**9a+9a'**), and (**9e+9e'**) (0.2 mmol) in THF (10 mL) was added NH_4OH (2 mmol) in THF (10 mL) at 0 °C and the reaction mixture was stirred for 2 h at 0 °C. THF was removed in vacuo, and the solid formed was dissolved in EtOAc (50 mL), washed with 5% Na_2CO_3 (3×15 mL), dried over MgSO_4 . Removing solvents under reduced pressure gave an oil, which was recrystallized from EtOAc/hexanes to give **10a–l**, (**10a+10a'**), and (**10e+10e'**).

4.5.1. Benzyl N-[(1S)-2-[(1S)-2-amino-1-methyl-2-oxoethyl]-amino-1-(1H-indol-2-ylmethyl)-2-oxoethyl]carbamate (Cbz-L-Trp-L-Ala-NH₂, **10a)**

Yield: 79%; white microcrystals; mp 168–170 °C; $[\alpha]_D^{23}$ –9.5 (c 1.42, DMF). ¹H NMR: δ =10.86 (s, 1H), 8.09 (d, *J*=7.4 Hz, 1H), 7.68 (d, *J*=7.7 Hz, 1H), 7.47 (d, *J*=8.1 Hz, 1H), 7.39–7.27 (m, 7H), 7.20 (s, 1H), 7.13–7.08 (m, 2H), 7.03–6.99 (m, 1H), 4.99 (s, 2H), 4.36–4.25 (m, 2H), 3.18 (dd, *J*=14.6, 4.0 Hz, 1H), 2.77 (dd, *J*=14.3, 9.9 Hz, 1H), 1.26 (d, *J*=7.0 Hz, 3H). ¹³C NMR: δ =174.2, 171.4, 155.9, 137.0, 136.1, 128.3, 127.7, 127.4, 127.3, 123.9, 120.9, 118.6, 118.2, 111.3, 110.2, 65.3, 55.6, 48.0, 27.7, 18.5. IR: $\nu_{\max}$ 3250, 3090, 3045, 2925, 1705, 1640, 1530, 1515, 1490, 1440 cm^{–1}. Anal. Calcd for C₂₂H₂₄N₄O₄: C, 64.69; H, 5.92; N, 13.72. Found: C, 64.95; H, 6.00; N, 13.32.

4.5.2. Benzyl N-[(1S)-2-[(1S)-2-amino-1-(1H-indol-2-ylmethyl)-2-oxoethyl]amino-1-(1H-indol-2-ylmethyl)-2-oxoethyl]carbamate (Cbz-L-Trp-L-Trp-NH₂, **10b)**

Yield: 96%; white microcrystals; mp 192–194 °C; $[\alpha]_D^{23}$ –36.6 (c 1.68, DMF). ¹H NMR: δ =10.84 (s, 1H), 10.81 (s, 1H), 8.03 (d, *J*=7.7 Hz, 1H), 7.61 (d, *J*=7.7 Hz, 2H), 7.44–7.44 (m, 2H), 7.34–7.27 (m, 5H), 7.24–7.22 (m, 2H), 7.16 (s, 1H), 7.11–7.08 (m, 2H), 7.06–7.03 (m, 2H), 6.99–6.94 (m, 2H), 4.94 (s, 2H), 4.56–4.52 (m, 1H), 4.33–4.28 (m, 1H), 3.18–2.99 (m, 3H), 2.88 (d, *J*=14.2, 10.2 Hz, 1H). ¹³C NMR: δ =173.4, 171.7, 156.0, 137.1, 136.2, 136.2, 128.5, 127.8, 127.6, 127.4, 123.9, 123.7, 121.0, 118.7, 118.4, 111.5, 111.4, 110.4, 110.1, 65.5, 55.9, 53.3, 28.0, 27.8. Anal. Calcd for C₃₀H₂₉N₅O₄: C, 68.82; H, 5.58; N, 13.38. Found: C, 68.65; H, 5.62; N, 13.40.

4.5.3. Benzyl N-[(1S)-2-[(2-amino-2-oxoethyl)amino]-1-(1H-indol-2-ylmethyl)-2-oxoethyl]carbamate (Cbz-L-Trp-Gly-NH₂, **10c)**

Yield: 86%; white microcrystals; mp 151–155 °C; $[\alpha]_D^{23}$ –26.4 (c 1.82, DMF). ¹H NMR: δ =10.86 (s, 1H), 8.32 (t, *J*=5.6 Hz, 1H), 7.66 (d, *J*=7.7 Hz, 1H), 7.54 (d, *J*=7.8 Hz, 1H), 7.39–7.35 (m, 3H), 7.32–7.28 (m, 3H), 7.23–7.21 (m, 2H), 7.16–7.08 (m, 2H), 7.01 (t, *J*=7.4 Hz, 1H), 4.99 (s, 2H), 4.36–4.29 (m, 1H), 3.70 (dd, *J*=12.4, 5.7 Hz, 2H), 3.19 (dd, *J*=14.6, 4.4 Hz, 1H), 2.97 (dd, *J*=14.6, 9.7 Hz, 1H). ¹³C NMR: δ =172.2, 171.0, 156.2, 137.1, 136.2, 128.5, 127.9, 127.7, 127.4, 124.0, 121.0, 118.6, 118.4, 111.5, 110.3, 65.5, 55.8, 42.2, 27.7. Anal. Calcd for C₂₁H₂₂N₄O₄: C, 63.95; H, 5.62; N, 14.20. Found: C, 64.10; H, 5.68; N, 13.82.

4.5.4. Benzyl N-[(1S)-2-[(1S)-2-amino-1-(1H-indol-2-ylmethyl)-2-oxoethyl]amino-1-methyl-2-oxoethyl]carbamate (Cbz-L-Ala-L-Trp-NH₂, **10d)**

Yield: 77%; white microcrystals; mp 201–202 °C; $[\alpha]_D^{23}$ –7.6 (c 1.62, DMF). ¹H NMR: δ =10.86 (s, 1H), 7.88 (d, *J*=8.0 Hz, 1H), 7.62 (d, *J*=7.7 Hz, 1H), 7.53 (d, *J*=7.3 Hz, 1H), 7.43–7.34 (m, 7H), 7.16–7.11 (m, 2H), 7.09–7.02 (m, 1H), 7.00 (t, *J*=7.3 Hz, 1H), 5.05 (AB, *J*_{AB}=19.4, 12.6 Hz, 2H), 4.50 (dd, *J*=13.1, 7.4 Hz, 1H), 4.09–4.06 (m, 1H), 3.16 (dd, *J*=14.7, 5.1 Hz, 1H), 3.03 (dd, *J*=14.6, 8.0 Hz, 1H), 1.19 (d, *J*=7.1 Hz, 3H). ¹³C NMR: δ =173.2, 172.2, 155.8, 137.0, 136.0, 128.4, 127.8, 127.4, 123.5, 120.8, 118.5, 118.2, 111.2, 110.0, 65.5, 53.0, 50.3, 27.6, 18.0. Anal. Calcd for C₂₂H₂₄N₄O₄: C, 64.69; H, 5.92; N, 13.72. Found: C, 64.78; H, 5.99; N, 13.51.

4.5.5. Benzyl (4S)-5-amino-4-[(2S)-2-[(benzyloxy)carbonyl]-aminopropanoyl]amino]-5-oxopentanoate (Cbz-L-Ala-L-Glu(4-OBn)-NH₂, **10e)**

Yield: 98%; white microcrystals; mp 162–163 °C; $[\alpha]_D^{23}$ –3.0 (c 1.32, DMF). ¹H NMR: δ =7.90 (d, *J*=8.0 Hz, 1H), 7.53 (d, *J*=7.1 Hz, 1H), 7.36–7.34 (m, 11H), 7.13 (s, 1H), 5.09 (s, 2H), 5.01 (d, *J*=7.7 Hz, 2H), 4.23 (dd, *J*=13.2, 8.1 Hz, 1H), 4.05 (t, *J*=7.1 Hz, 1H), 2.37 (t, *J*=7.8 Hz, 2H), 2.09–2.00 (m, 1H), 1.84–1.77 (m, 1H), 2.40 (d, *J*=7.0 Hz, 3H). ¹³C NMR: δ =172.9, 172.4, 155.8, 137.0, 136.2, 128.4, 128.3, 128.0, 127.9, 127.7, 65.5, 65.4, 51.5, 50.2, 30.0, 27.3, 17.9. Anal. Calcd for C₂₃H₂₇N₃O₆: C, 62.57; H, 6.16; N, 9.52. Found: C, 62.57; H, 6.19; N, 9.45.

4.5.6. Benzyl (4S)-5-amino-4-[(2-[(benzyloxy)carbonyl]-aminopropanoyl]amino)-5-oxopentanoate (Cbz-DL-Ala-L-Glu(4-OBn)-NH₂, (10e**+**10e'**))**

Yield: 90%; white microcrystals; mp 138–139 °C; $[\alpha]_D^{23}$ –4.7 (c 1.33, DMF). ¹H NMR: δ =8.12–7.93 (m, 1H), 7.58 (t, *J*=6.2 Hz, 1H), 7.44–7.34 (m, 11H), 7.19 (d, *J*=15.5 Hz, 1H), 5.12 (s, 2H), 5.05 (s, 2H), 4.29–4.23 (m, 1H), 4.13–4.04 (m, 1H), 2.44–2.36 (m, 2H), 2.10–1.98 (m, 1H), 1.88–1.81 (m, 1H), 1.23 (dd, *J*=7.0, 1.2 Hz, 3H). ¹³C NMR: δ =172.9, 172.8, 172.6, 172.4, 172.3, 155.9, 155.8, 137.0, 136.9, 136.2, 128.4, 128.3, 128.0, 127.9, 127.8, 65.5, 51.4, 50.2, 30.0, 27.3, 27.0, 18.0. Anal. Calcd for C₂₃H₂₇N₃O₆: C, 62.57; H, 6.16; N, 9.52. Found: C, 62.42; H, 6.16; N, 9.65.

4.5.7. Benzyl N-[(1S)-2-[(2S)-2-(aminocarbonyl)tetrahydro-1H-pyrrol-1-yl]-1-methyl-2-oxoethyl]carbamate (Cbz-L-Ala-L-Pro-NH₂, **10f)**

Yield: 95%; white microcrystals; mp 161–162 °C; $[\alpha]_D^{23}$ –37.6 (c 1.88, DMF). ¹H NMR: δ =7.52 (d, *J*=7.4 Hz, 1H), 7.39–7.25 (m, 5H), 7.21 (br s, 1H), 6.89 (br s, 1H), 5.00 (s, 2H), 4.31 (t, *J*=7.0 Hz, 1H), 4.21 (dd, *J*=8.20, 3.5 Hz, 1H), 3.59–3.55 (m, 2H), 2.05–1.76 (m, 4H), 1.20 (d, *J*=6.7 Hz, 3H). ¹³C NMR: δ =173.6, 170.8, 155.7, 137.1, 128.4, 127.8, 127.7, 65.3, 59.5, 48.0, 46.5, 29.2, 24.5, 16.8. Anal. Calcd for C₁₆H₂₁N₃O₄: C, 60.18; H, 6.63; N, 13.16. Found: C, 60.00; H, 6.73; N, 12.96.

4.5.8. Benzyl N-[(1S)-1-[(1S)-2-amino-1-(1H-indol-2-ylmethyl)-2-oxoethyl]aminocarbonyl]-3-(methylsulfanyl)propyl]carbamate (Cbz-L-Met-L-Trp-NH₂, **10g)**

Yield: 91%; white microcrystals; mp 218–220 °C; $[\alpha]_D^{23}$ –12.7 (c 1.50, DMF). ¹H NMR: δ =10.8 (s, 1H), 7.90 (d, *J*=7.8 Hz, 1H), 7.59 (d, *J*=7.8 Hz, 1H), 7.54 (d, *J*=7.8 Hz, 1H), 7.43 (s, 1H), 7.35–7.27 (m, 6H), 7.13–7.03 (m, 3H), 6.96 (t, *J*=7.3 Hz, 1H), 5.02 (d, *J*=4.4 Hz, 2H), 4.52–4.45 (m, 1H), 4.08–4.04 (m, 1H), 3.13 (dd, *J*=15.0, 5.1 Hz, 1H), 3.00 (dd, *J*=14.9, 8.0 Hz, 1H), 2.39 (t, *J*=7.6 Hz, 2H), 1.99 (s, 3H), 1.84–1.73 (m, 2H). ¹³C NMR: δ =173.4, 171.3, 156.2, 137.1, 136.1, 128.5, 128.0, 127.9, 127.6, 123.6, 121.0, 118.7, 118.4, 111.4, 110.1, 65.7, 54.2, 53.2, 31.8, 29.7, 27.8, 14.7. Anal. Calcd for C₂₄H₂₈N₄O₄S: C, 61.52; H, 6.02; N, 11.96. Found: C, 61.69; H, 6.16; N, 11.68.

4.5.9. Benzyl N-[(1S)-2-[(1S)-1-(aminocarbonyl)-3-(methylsulfanyl)propyl]amino-1-benzyl-2-oxoethyl]carbamate (Cbz-L-Phe-L-Met-NH₂, **10h)**

Yield: 94%; white microcrystals; mp 208–210 °C; $[\alpha]_D^{23}$ –14.8 (c 0.92, DMF). ¹H NMR: δ =8.13 (d, *J*=8.1 Hz, 1H), 7.61 (d, *J*=8.4 Hz, 1H), 7.40–7.19 (m, 11H), 7.15 (s, 1H), 4.98 (s, 2H), 4.36–4.27 (m, 2H), 3.06 (dd, *J*=13.8, 4.1 Hz, 1H), 2.78 (dd, *J*=13.6, 10.6 Hz, 1H), 2.50–2.46 (m, 2H), 2.07 (s, 3H), 2.00–1.95 (m, 1H), 1.85–1.81 (m, 1H). ¹³C NMR: δ =173.1, 171.6, 156.1, 138.2, 137.1, 129.4, 128.5, 128.2, 127.9, 127.6, 126.4, 65.4, 56.4, 51.9, 38.0, 32.1, 29.7, 14.8. Anal. Calcd for C₂₂H₂₇N₃O₄S: C, 61.52; H, 6.34; N, 9.78. Found: C, 61.14; H, 6.39; N, 9.50.

4.5.10. Benzyl (4S)-5-amino-4-[(2S)-2-[(benzyloxy)carbonyl]-amino-3-phenylpropanoyl]amino]-5-oxopentanoate (Cbz-L-Phe-L-Glu(4-OBn)-NH₂, **10i)**

Yield: 96%; white microcrystals; mp 205–207 °C; $[\alpha]_D^{23}$ –10.6 (c 1.1, DMF). ¹H NMR: δ =8.10 (d, *J*=8.0 Hz, 1H), 7.60 (d, *J*=8.4 Hz, 1H), 7.41–7.38 (m, 5H), 7.37–7.22 (m, 11H), 7.16 (s, 1H), 5.13 (s, 2H), 4.96 (s, 2H), 4.34–4.24 (m, 2H), 3.05 (dd, *J*=13.9, 4.0 Hz, 1H), 2.78 (dd, *J*=13.6, 10.7 Hz, 1H), 2.41 (t, *J*=8.0 Hz, 2H), 2.06–1.79 (m, 2H). ¹³C NMR: δ =172.9, 172.4, 171.6, 156.0, 138.2, 137.1, 136.4, 129.3, 128.6, 128.5, 128.2, 128.2, 128.1, 127.9, 127.6, 126.4, 65.7, 65.4, 56.4, 51.8, 37.3, 30.2, 27.5. Anal. Calcd for C₂₉H₃₁N₃O₆: C, 66.99; H, 6.04; N, 8.12. Found: C, 67.00; H, 6.10; N, 7.92.

4.5.11. Benzyl (3S)-4-amino-3-[(2S)-2-[(benzyloxy)carbonyl]-amino-3-phenylpropanoyl]amino]-4-oxobutanoate (Cbz-L-Phe-L-Asp(3-OBn)-NH₂, **10j)**

Yield: 94%; white microcrystals; mp 163–165 °C; $[\alpha]_D^{23}$ –28.6 (c 1.61, DMF). ¹H NMR: δ =8.39 (d, *J*=8.0 Hz, 1H), 7.61 (d, *J*=8.1 Hz, 1H), 7.40–7.25 (m, 17H), 5.13 (s, 2H), 4.97 (s, 2H), 4.61 (q, *J*=7.7 Hz, 1H), 4.34–4.30 (m, 1H), 3.04 (d, *J*=9.9 Hz, 1H), 2.91–2.64 (m, 3H). ¹³C NMR: δ =172.1, 171.6, 170.3, 156.1, 138.2, 137.1, 136.2, 129.4, 128.6, 128.5, 128.2, 128.1, 128.1, 127.9, 127.6, 126.4, 65.9, 65.4, 56.4, 49.5, 37.4, 36.4. Anal. Calcd for C₂₈H₂₉N₃O₆: C, 66.79; H, 5.80; N, 8.34. Found: C, 66.55; H, 5.82; N, 8.20.

4.5.12. Benzyl N-((5S)-6-amino-5-[(2S)-2-[(benzyloxy)carbonyl]-amino-3-phenylpropanoyl]amino)-6-oxohexylcarbamate (Cbz-L-Phe-L-N⁶-Lys-NH₂, **10k)**

Yield: 92%; white microcrystals; mp 193–195 °C; $[\alpha]_D^{23}$ –8.5 (c 1.68, DMF). ¹H NMR: δ =8.05 (d, *J*=8.0 Hz, 1H), 7.59 (d, *J*=8.5 Hz, 1H), 7.38–7.26 (m, 17H), 7.09 (s, 1H), 5.04 (s, 2H), 4.98 (s, 2H), 4.34–4.20 (m, 2H), 3.09–2.98 (m, 3H), 2.78 (t, *J*=12.6 Hz, 1H), 1.74–1.52 (m, 2H), 1.46–1.40 (m, 2H), 1.34–1.27 (m, 2H). ¹³C NMR: δ =173.6, 171.5, 156.2, 156.0, 138.3, 137.4, 137.2, 129.4, 128.5, 128.4, 128.2, 127.9, 127.8, 127.6, 127.5, 126.4, 65.4, 65.3, 56.4, 52.5, 40.5, 37.5, 32.7, 29.3, 22.7. Anal. Calcd for C₃₁H₃₆N₄O₆: C, 66.41; H, 6.47; N, 9.99. Found: C, 66.20; H, 6.37; N, 9.70.

4.5.13. Benzyl (3S)-4-amino-3-[(2R)-2-[(benzyloxy)carbonyl]-amino-3-methylbutanoyl]amino]-4-oxobutanoate (Cbz-D-Val-L-Asp(3-OBn)-NH₂, **10l)**

Yield: 88%; white microcrystals; mp 195–195 °C; $[\alpha]_D^{23}$ –33.0 (c 1.70, DMF). ¹H NMR: δ =8.46 (d, *J*=8.4 Hz, 1H), 7.49 (d, *J*=7.4 Hz, 1H), 7.40–7.23 (m, 12H), 5.06 (d, *J*=2.1 Hz, 2H), 5.02 (s, 2H), 4.68–4.61 (m, 1H), 3.75 (t, *J*=7.4 Hz, 1H), 2.89 (dd, *J*=16.1, 4.8 Hz, 1H), 2.60 (dd, *J*=16.2, 9.1 Hz, 1H), 1.91–1.85 (m, 1H), 0.84–0.78 (m, 6H). ¹³C NMR: δ =172.2, 171.4, 170.3, 156.5, 136.9, 136.0, 128.4, 128.0, 127.9, 127.9, 65.7, 65.5, 60.8, 49.2, 36.1, 29.6, 19.0, 18.6. Anal. Calcd for C₂₄H₂₉N₃O₆: C, 63.28; H, 6.42; N, 9.22. Found: C, 63.18; H, 6.48; N, 9.03.

4.6. Tripeptide primary amides **13: general procedure**

A mixture of *N*-unprotected amino amide **12a,b** (0.2 mmol) and *N*-protected dipeptidylbenzotriazoles **9a–I**, (**9a+9a'**), and (**9e+9e'**) (0.2 mmol) in acetonitrile (10 mL) was stirred for 6 h at –15 °C. Acetonitrile was removed in vacuo, and the solid formed was dissolved in EtOAc (50 mL), washed with 5% Na₂CO₃ (3×15 mL), and dried over MgSO₄. Removing solvents under reduced pressure gave oil, which was recrystallized from EtOAc/hexanes to give **13a,b** and (**13a+13a'**).

4.6.1. Benzyl N-((1S)-1-[(1S)-2-[(1S)-1-(aminocarbonyl)-3-methylsulfanyl]propyl]amino-1-methyl-2-oxoethylamino)-1-(1H-indol-2-ylmethyl)-2-oxoethyl]carbamate (Cbz-L-Trp-L-Ala-L-Met-NH₂, **13a)**

Yield: 82%; white microcrystals; mp 222–224 °C. ¹H NMR: δ =10.86 (s, 1H), 8.33–8.28 (m, 1H), 7.92–7.90 (m, 1H), 7.71 (d, *J*=8.0 Hz, 1H), 7.51 (d, *J*=8.1 Hz, 1H), 7.38–7.27 (m, 7H), 7.21–7.17 (m, 2H), 7.10 (t, *J*=7.4 Hz, 1H), 7.01 (t, *J*=7.3 Hz, 1H), 5.00–4.97 (m, 2H), 4.37–4.24 (m, 3H), 3.18–3.14 (m, 1H), 2.98–2.90 (m, 1H), 2.51–2.39 (m, 2H), 2.06 (s, 3H), 2.00–1.94 (m, 1H), 1.88–1.76 (m, 1H), 1.28–1.12 (m, 3H). ¹³C NMR: δ =173.1, 172.2, 172.1, 156.1, 137.1, 136.2, 128.5, 127.8, 127.7, 127.6, 127.4, 124.1, 121.0, 118.8, 118.3, 111.4, 110.3, 65.4, 55.6, 51.8, 48.7, 32.1, 29.7, 17.9, 14.8. Anal. Calcd for C₂₇H₃₃N₅O₅S: C, 60.09; H, 6.16; N, 12.98. Found: C, 59.78; H, 6.25; N, 12.68.

4.6.2. Benzyl N-((1S)-2-[(2-amino-2-oxoethyl)amino]-1-methyl-2-oxoethylamino)-1-(1H-indol-2-ylmethyl)-2-oxoethyl]carbamate (Cbz-L-Trp-DL-Ala-Gly-NH₂, (13a+13a'**))**

Yield: 86%; white microcrystals; mp 164–166 °C. ¹H NMR: δ =10.84 (s, 1H), 8.36–8.28 (m, 1H), 8.06–8.00 (m, 1H), 7.68–7.60 (m, 1H), 7.53–7.46 (m, 1H), 7.35–7.24 (m, 7H), 7.17–7.13 (m, 2H), 7.06 (t, *J*=7.4 Hz, 1H), 6.97 (t, *J*=7.2 Hz, 1H), 4.99 (s, 2H), 4.29–4.23 (m, 2H), 3.70–3.60 (m, 2H), 3.16–3.06 (m, 1H), 2.98–2.87 (m, 1H), 1.24–1.12 (m, 3H). ¹³C NMR: δ =172.3, 172.2, 172.0, 170.9, 156.1, 156.0, 136.9, 137.0, 136.9, 136.1, 128.3, 127.8, 127.7, 127.5, 127.3, 124.0, 120.8, 118.6, 118.5, 118.2, 111.3, 110.1, 109.8, 65.5, 65.3, 55.7, 55.5, 48.6, 41.9, 27.6, 17.8, 17.7. HRMS calcd for C₂₄H₂₇N₅O₅ [M+H]⁺ 466.2085, found 466.2063.

4.6.3. Benzyl N-((1S)-2-[(1S)-2-[(2-amino-2-oxoethyl)amino]-1-(1H-indol-2-ylmethyl)-2-oxoethyl]amino)-1-(1H-indol-2-ylmethyl)-2-oxoethyl]carbamate (Cbz-L-Trp-L-Trp-Gly-NH₂, **13b)**

Yield: 86%; white microcrystals; mp 220–221 °C. ¹H NMR: δ =10.90 (s, 1H), 10.85 (s, 1H), 8.30–8.23 (m, 2H), 7.68–7.61 (m, 2H), 7.50 (d, *J*=8.2 Hz, 1H), 7.38–7.32 (m, 5H), 7.29–7.23 (m, 4H), 7.18–7.15 (m, 2H), 7.10 (t, *J*=7.4 Hz, 2H), 7.04–6.98 (m, 2H), 4.98 (s, 2H), 4.59 (q, *J*=12.9 Hz, 1H), 4.38–4.30 (m, 1H), 3.74 (dd, *J*=16.8, 6.0 Hz, 1H), 3.60 (dd, *J*=16.8, 5.3 Hz, 1H), 3.24 (dd, *J*=16.7, 5.0 Hz, 1H), 3.14–3.04 (m, 2H), 2.91 (dd, *J*=14.6, 10.2 Hz, 1H). ¹³C NMR: δ =172.1, 171.5, 170.9, 155.9, 136.9, 136.1, 128.3, 127.7, 127.5, 127.4, 127.3, 123.8, 123.7, 120.9, 118.6, 118.4, 118.2, 111.3, 110.2, 109.9, 65.4, 55.6, 53.8, 42.1, 27.6, 27.5. Anal. Calcd for C₃₂H₃₂N₆O₅: C, 66.19; H, 5.55; N, 14.47. Found: C, 65.84; H, 5.61; N, 14.28.

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References and notes

- (a) Kreil, G. *Methods Enzymol.* **1984**, 106, 218; (b) Pavia, M. R.; Sawyer, T. K.; Moos, W. H. *Bioorg. Med. Chem. Lett.* **1993**, 3, 387.
- Castillo, M. J.; Nakajima, K.; Zimmerman, M.; Powers, K. *Anal. Biochem.* **1979**, 99, 53.
- Davies, M.; Bradley, M. *Tetrahedron* **1999**, 55, 4733.
- (a) Fisher, J. M.; Scheller, R. H. *J. Biol. Chem.* **1988**, 263, 16515; (b) Mortensen, U. H.; Raaschou-Nielsen, M.; Breddam, K. *J. Biol. Chem.* **1994**, 269, 15528.
- Borthwick, A. D.; Davies, D. E.; Exall, A. M.; Hatley, R. J. D.; Hughes, J. A.; Irving, W. R.; Livermore, D. G.; Sollis, S. L.; Nerozzi, F.; Valko, K. L.; Allen, J.; Perren, M.; Shabbir, S. S.; Woollard, P. M.; Price, M. A. *J. Med. Chem.* **2006**, 49, 4159.
- Krebs, L. J.; Wang, X.; Pudavar, H. E.; Bergey, E. J.; Schally, A. V.; Nagy, A.; Prasad, P. N.; Liebow, C. *Cancer Res.* **2000**, 60, 4194.
- Fukuhara, S.; Mukai, H.; Kako, K.; Nakayama, K.; Munekata, E. *J. Neurochem.* **1996**, 67, 1282.
- Glass, J. D.; Schwartz, I. L.; Walter, R. *Proc. Natl. Acad. Sci. U.S.A.* **1969**, 63, 1426.
- (a) Dias, L. C.; Ferreira, A. A.; Diaz, G. *Synlett* **2002**, 1845; (b) De Clercq, E. *Pure Appl. Chem.* **2001**, 73, 55; (c) Ghosh, A. K.; Shin, D.; Mathivanan, P. *Chem. Commun.* **1999**, 1025.
- Shearman, M. S.; Behr, D.; Clarke, E. E.; Lewis, H. D.; Harrison, T.; Hunt, P.; Nadin, A.; Smith, A. L.; Stevenson, G.; Castro, J. L. *Biochemistry* **2000**, 39, 8698.
- Brinker, A.; Weber, E.; Stoll, D.; Voigt, J.; Mueller, A.; Sewald, N.; Jung, G.; Wiesmueller, K.-H.; Bohley, P. *Eur. J. Biochem.* **2000**, 267, 5085.
- Brix, K.; Lemansky, P.; Herzog, V. *Endocrinology* **1996**, 137, 1963.
- (a) Fletcher, M. D.; Campbell, M. M. *Chem. Rev.* **1998**, 98, 763; (b) Tierney, D. L.; Huang, H.; Martasek, P.; Roman, L. J.; Silverman, R. B.; Hoffman, B. M. *J. Am. Chem. Soc.* **2000**, 122, 7869.
- Huang, H.; Martasek, P.; Roman, L. J.; Masters, B. S. S.; Silverman, R. B. *J. Med. Chem.* **1999**, 42, 3147.
- Choi, D. W.; Rothman, S. M. *Annu. Rev. Neurosci.* **1990**, 13, 171.
- Olesen, J.; Thomsen, L. L.; Iversen, H. *Trends Pharmacol. Sci.* **1994**, 15, 149.
- Dorheim, M. A.; Tracey, W. R.; Pollock, J. S.; Grammas, P. *Biochem. Biophys. Res. Commun.* **1994**, 205, 659.
- (a) Katritzky, A. R.; Suzuki, K.; Wang, Z. *Synlett* **2005**, 1656; (b) Katritzky, A. R.; Yang, H.; Zhang, S.; Wang, M. *ARKIVOC* **2002**, xi, 39; (c) Katritzky, A. R.; Wang, M.; Yang, H.; Zhang, S.; Akhmedov, N. G. *ARKIVOC* **2002**, vii, 134; (d) Katritzky, A. R.; Abdel-Fattah, A. A.; Akhmedova, R. *ARKIVOC* **2005**, vi, 239.
- (a) Katritzky, A. R.; Abdel-Fattah, A. A.; Gromova, A.; Rachel Witte, R.; Peter Steel, P. *J. Org. Chem.* **2005**, 70, 9211; (b) Katritzky, A. R.; Abdel-Fattah, A. A. A.

- Wang, M. *J. Org. Chem.* **2003**, 68, 1443; (c) Katritzky, A. R.; Abdel-Fattah, A. A. A.; Wang, M. *J. Org. Chem.* **2003**, 68, 4932.
20. (a) Katritzky, A. R.; Pastor, A. *J. Org. Chem.* **2000**, 65, 3679; (b) Katritzky, A. R.; Pastor, A.; Voronkov, M. V. *J. Heterocycl. Chem.* **1999**, 36, 777.
21. (a) Katritzky, A. R.; Angrish, P. *Steroids* **2006**, 71, 660; (b) Katritzky, A. R.; Suzuki, K.; Singh, S. *Synthesis* **2004**, 2645; (c) Katritzky, A. R.; Angrish, P.; Hur, D.; Suzuki, K. *Synthesis* **2005**, 397; (d) Katritzky, A. R.; Angrish, P.; Suzuki, K. *Synthesis* **2006**, 411; (e) Katritzky, A. R.; Todadze, E.; Cusido, J.; Angrish, P.; Shestopalov, A. A. *Chem. Biol. Drug Des.* **2006**, 68, 37; (f) Katritzky, A. R.; Todadze, E.; Shestopalov, A. A.; Cusido, J.; Angrish, P. *Chem. Biol. Drug Des.* **2006**, 68, 42; (g) Katritzky, A. R.; Meher, G.; Angrish, P. *Chem. Biol. Drug Des.* **2006**, 68, 326; (h) Katritzky, A. R.; Todadze, E.; Angrish, P.; Draghici, B. *J. Org. Chem.* **2007**, 72, 5794.
22. Compound numbers written within brackets represent a diastereomeric mixtures or racemates; compound numbers without brackets represent enantiomers.
23. Carpino, L. A.; Han, G. Y. *J. Org. Chem.* **1972**, 37, 3404.
24. Vogel, A. *Vogel's textbook of Practical Organic Chemistry*; Langman Scientific & Technical and Wiley: New York, NY, 1989.
25. March, J. *Advanced Organic Chemistry: Reactions, Mechanisms, and Structure*; John Wiley & Sons: New York, NY, 1992.
26. Bodanszky, M. *The Practice of Peptide Synthesis*; Springer: Berlin, 1984; Galaverra, G.; Corradini, R.; Dossena, A.; Marchelli, R. *Int. J. Pept. Protein Res.* **1993**, 42, 53.
27. Wang, W.; McMurray, J. S. *Tetrahedron Lett.* **1999**, 40, 2501.
28. Carpino, L. A. *J. Am. Chem. Soc.* **1993**, 115, 4397.
29. Ananda, K.; Vasanthakumar, G. R.; Babu, V. V. S. *Protein Pept. Lett.* **2001**, 8, 45.
30. Nozaki, S.; Muramatsu, I. *Bull. Chem. Soc. Jpn.* **1988**, 61, 2647.
31. (a) Rink, H. *Tetrahedron Lett.* **1987**, 28, 3787; (b) Bernatowicz, M. S.; Daniels, S. B.; Koster, H. *Tetrahedron Lett.* **1989**, 30, 4645.
32. Atherton, E.; Logan, C. J.; Sheppard, R. C. *J. Chem. Soc., Perkin Trans. 1* **1981**, 538.
33. Bray, A. M.; Jhingran, A. G.; Valerio, R. M.; Maeji, N. J. *J. Org. Chem.* **1994**, 59, 2197.
34. Brown, E.; Sheppard, R. C.; Williams, B. J. *J. Chem. Soc., Perkin Trans. 1* **1983**, 75.
35. Cros, E.; Planas, M.; Mejias, X.; Bardaji, E. *Tetrahedron Lett.* **2001**, 42, 6105.
36. Ramer, S. E.; Cheng, H.; Vederas, J. C. *Pure Appl. Chem.* **1989**, 61, 489.
37. (a) Miyazawa, T.; Nakajo, S.; Nishikawa, M.; Imagawa, K.; Yanagihara, R.; Yamada, T. *J. Chem. Soc., Perkin Trans. 1* **1996**, 2867; (b) Rival, S.; Besson, C.; Saulnier, J.; Wallach, J. J. *Peptide Res.* **1999**, 53, 170; (c) Matsumoto, K.; Davis, B. G.; Jones, B. *Chem.—Eur. J.* **2002**, 8, 4129; (d) Wehofsky, N.; Koglin, N.; Thus, S.; Bordusa, F. *J. Am. Chem. Soc.* **2003**, 125, 6126; (e) Abdus Salam, S. M.; Kagawa, K.-I.; Kawashiro, K. *Biotechnol. Lett.* **2005**, 27, 1199.
38. Miyazawa, T.; Hiramatsu, M.; Murashima, T.; Yamada, T. *Biocatal. Biotransform.* **2003**, 21, 93.